Nikon

Nikon 100th anniversary

C2+

Confocal Microscope

Simple and Robust

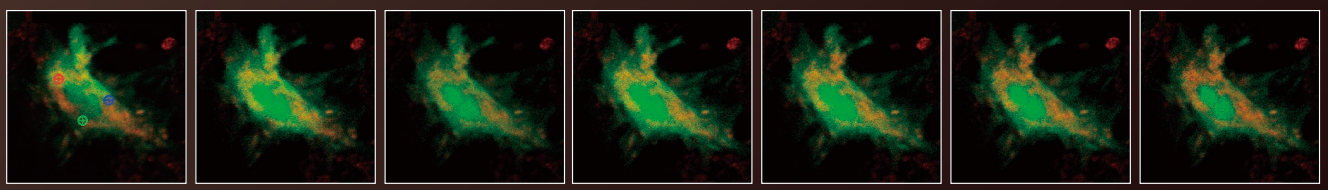
An essential microscopy laboratory instrument…

The C2+ confocal microscope is a basic model within the family of Nikon confocal products. The C2+ is designed as an essential microscopy tool for the laboratory, providing powerful and robust imaging capabilities. The high-efficiency scan heads and detectors, coupled with Nikon's unrivaled optics, provide superior confocal images. The high-speed galvano scanners, operating at rates of up to 100 fps*, enable even the fast-beating motion of cardiac muscles to be captured with precision. The system also provides simultaneous acquisition of three fluorescent channels plus DIC in a single scan. For research that requires spectral imaging capabilities, Nikon's C2si+ system provides dedicated spectral detector units in addition to the standard fluorescence detector units. The C2si+ system allows high-precision and high-speed 32-channnel spectral imaging or high-sensitivity spectral imaging. Built on a reputation of incredible stability and operational simplicity, coupled with superior optical technologies, the C2+ confocal system is the essential laboratory tool.

* with 8x zoom or larger

High-speed acquisition of high-definition images

Galvanometer-based high-speed scanning enables confocal imaging of fast, dynamic events in live cells, such as the beating motion of cardiac muscle cells. In traditional confocal systems, fast bi-directional scanning can lead to pixel shifts. However, the C2+ confocal system's pixel shift correction mechanism ensures acquisition of the highest quality images even with fast bi-directional scanning.



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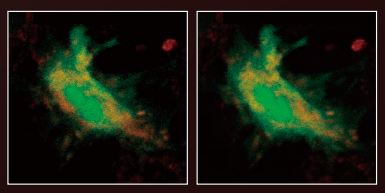

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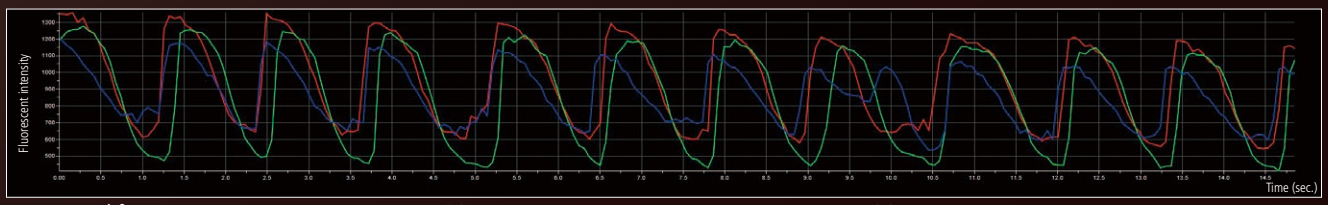

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Rat primary cardiac muscle cells labeled with Fluo-8 AM and MitoTracker® Orange are stimulated with histamine. The graph shows the intensity change of each ROI (red, green and blue) in the first image shown above. Scanning speed: 15 fps

Intuitive operation

The superior operability and diverse analysis capabilities of the NIS-Elements imaging software satisfy both beginners and experienced confocal users. In addition, NIS-Elements allows for intuitive operation not only of Nikon microscopes but also of third party peripheral devices for a broad range of experiments.

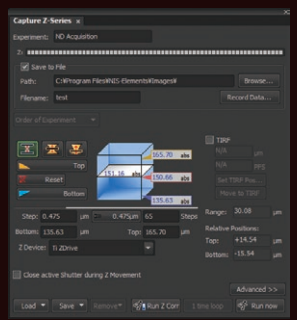
Multimode capability

Various imaging methods, such as confocal, widefield, TIRF, photoactivation, as well as processing, analysis and presentation of acquired images, are available in one software package. Users can easily learn how to control different imaging systems with a common interface and workflow.

Setting: Easy-to-recognize display for setting lasers, detectors, etc.

Simple GUI: Simple display of fundamental image acquisition settings

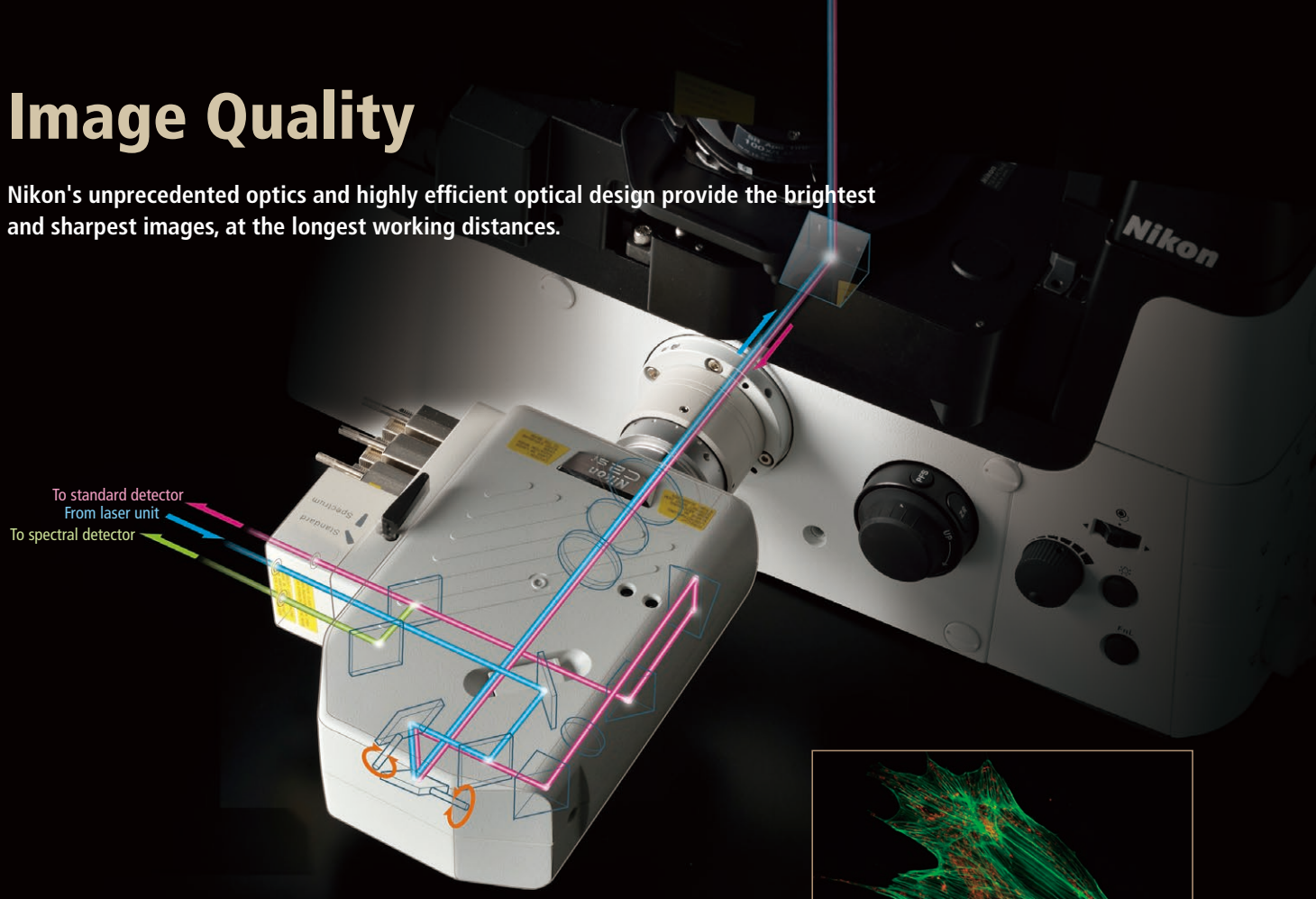
Scanning parameter settings

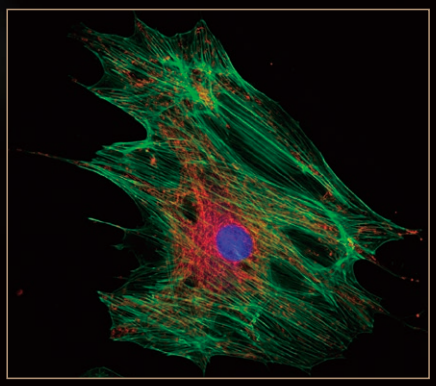


Intuitive settings for Z series parameters

Image Quality

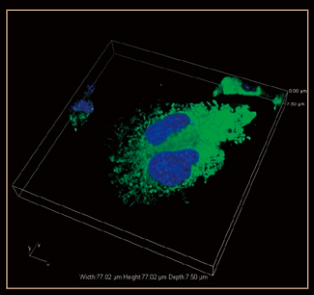
Nikon's unprecedented optics and highly efficient optical design provide the brightest and sharpest images, at the longest working distances.

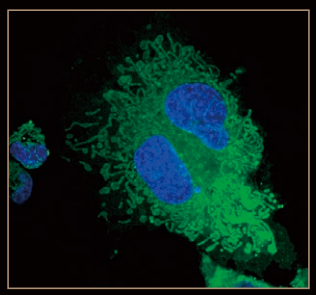


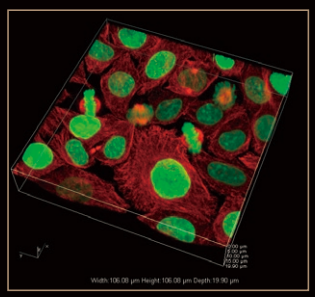


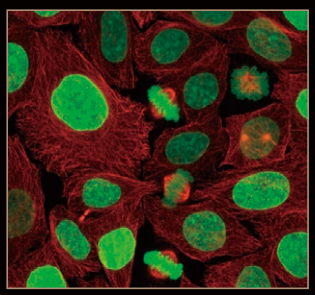
High-efficiency scan heads and detectors

With the convenient, small scan head size, the C2+ can be used with various types of Nikon microscope. The C2+ employs high precision mirrors and optically superior circular pinholes, and separates the detectors to isolate sources of heat and noise, enabling low-noise, high-contrast and high-quality confocal imaging. The newly developed scanner driving system and Nikon's unique image correction technique allow 8 fps (512 x 512 pixels) and 100 fps (512 x 32 pixels) high-speed imaging.





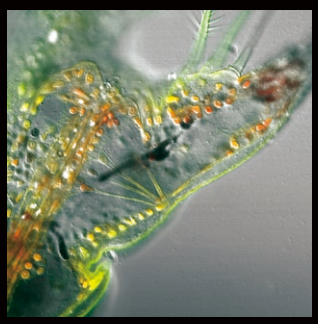




High-definition diascopic DIC images

The C2+ can acquire simultaneous three-channel fluorescence or simultaneous three-channel and diascopic DIC observation. High-quality DIC images and fluorescence images can be superimposed to aid in morphological analysis.

DIC image



Overlay of DIC and fluorescence images

High-performance objectives for confocal imaging

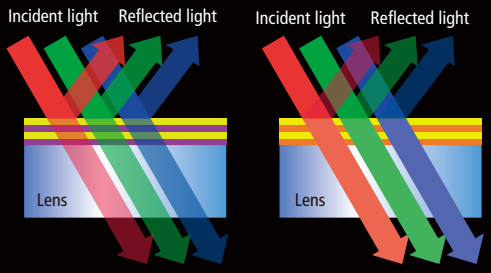
CFI Apochromat Lambda S Series

These high-numerical-aperture (NA) objectives provide chromatic aberration correction over a wavelength ranging from ultraviolet to infrared and are ideal for multicolor confocal imaging. In particular, the LWD Lambda S 40XC WI lens has an extremely wide chromatic aberration correction range from 405 nm to near-IR. The transmission property of these lenses is enhanced through the use of Nikon's exclusive Nano Crystal Coat technology.

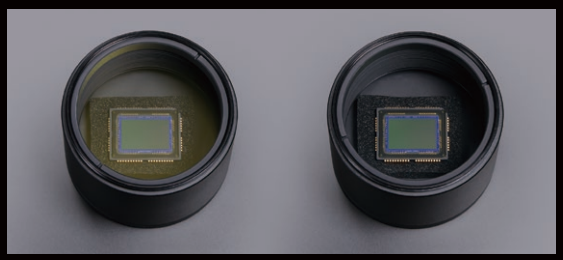
CFI Apochromat LWD Lambda S 20XC WI
CFI Apochromat LWD Lambda S 40XC WI
CFI Apochromat Lambda S 40XC WI
CFI Apochromat Lambda S 60X Oil

Nano Crystal Coat technology

With its origins in Nikon's semiconductor manufacturing technology, Nano Crystal Coat is an anti-reflective coating that assimilates ultra-fine crystallized particles of nanometer size. With particles arranged in a spongy construction with uniform spaces between them, this coarse structure enables lower refractive indices, facilitating the passage of light through the lens. These crystallized particles eliminate reflections inside the lens throughout the spectrum of visible light waves in ways that far exceed the limits of conventional anti-reflective coating systems.



Conventional multi-layer coating
Nano Crystal Coat



Conventional multi-layer coating
Nano Crystal Coat

CFI Apochromat TIRF 60XC Oil/1.49 (left)
CFI Apochromat TIRF 100XC Oil/1.49 (right)

CFI Apochromat TIRF Series

These objectives boast an unprecedented NA of 1.49 (using a standard coverslip and immersion oil), the highest resolution among Nikon objectives. Correction collars enable optimization of point spread functions for varying imaging temperatures, ensuring highest quality confocal images whether you are imaging at room temperature or 37 degrees Celsius.

Recommended objective lenses

CFI Plan Apochromat Lambda 10X	NA 0.45, W.D. 4.00 mm	N
CFI Plan Apochromat VC 20X	NA 0.75, W.D. 1.00 mm	
CFI Apochromat LWD Lambda S 20XC WI	NA 0.95, W.D. 0.95 mm	N
CFI Plan Apochromat Lambda 40XC	NA 0.95, W.D. 0.21 mm	N
CFI Apochromat Lambda S 40XC WI	NA 1.25, W.D. 0.20-0.16	N
CFI Apochromat LWD Lambda S 40XC WI	NA 1.15, W.D. 0.60 mm	N
CFI Apochromat Lambda S 60X Oil	NA 1.40, W.D. 0.14 mm	N
CFI Plan Apochromat VC 60XC WI	NA 1.20, W.D. 0.29 mm	
CFI Apochromat TIRF 60XC Oil	NA 1.49, W.D. 0.12 mm	
CFI Plan Apochromat IR 60XC WI	NA 1.27, W.D. 0.18-0.16 mm	N
CFI Apochromat TIRF 100XC Oil	NA 1.49, W.D. 0.12 mm	

N : Nano Crystal Coat-deposited

CFI Plan Apochromat Lambda 10X

CFI Plan Apochromat VC 20X

CFI Plan Apochromat Lambda 40XC

CFI Plan Apochromat VC 60XC WI

CFI Plan Apochromat IR 60XC WI

Enhanced Spectral Imaging

The C2si+ provides 2 options for spectral imaging: a fast and accurate 32-channel spectral detector unit and the newly-developed high-sensitivity GaAsP detector unit. The C2si+ allows not only clean separation of overlapping spectra of fluorescent labels in multi-stained specimens, but also unique user-definable "Virtual Filter" emission filter mode.

C2-DUS Spectral Detector Unit

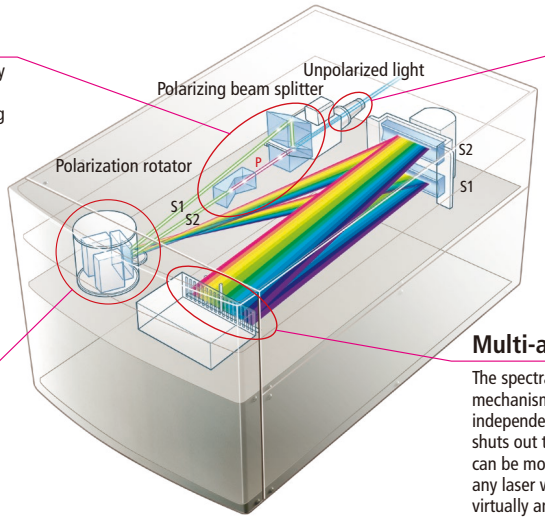
The C2-DUS allows for acquisition of 32 channels of fluorescence spectra over a 320 nm wavelength range with a single scan. By precisely unmixing the overlapping spectra at a high wavelength resolution of at least 2.5 nm, the C2-DUS facilitates the acquisition of detailed data.

DEES system

High diffraction efficiency is achieved by matching the polarization direction of light entering a grating to the polarizing light beam S.

Optical fiber

The wavelength resolution is independent of pinhole diameter.



Multiple gratings

Wavelength resolution can be varied between 2.5/5/10 nm with three gratings. Each position is precisely controlled for high-wavelength reproducibility.

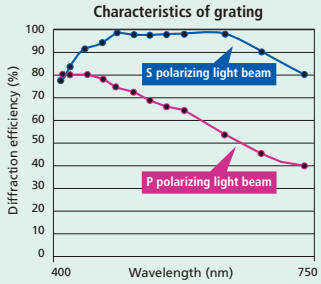
Multi-anode PMT

The spectral imaging detector utilizes a laser shielding mechanism. Coupled with a wavelength resolution independent of pinhole diameter, this mechanism successfully shuts out the reflected laser beam. The blocking mechanism can be moved freely with software, allowing users to block any laser wavelength, making the C2si+ compatible with virtually any laser selection.

High-quality spectral data acquisition

Diffraction Efficiency Enhancement System (DEES)

With the DEES, unpolarized fluorescence light emitted by the specimen is separated into two polarizing light beams P and S by a polarizing beam splitter. P is converted by a polarization rotator into S, which has higher diffraction efficiency than P, achieving vastly increased overall diffraction efficiency.

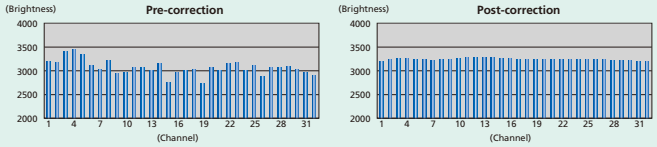


High-efficiency fluorescence transmission technology

The ends of the fluorescence fibers and detector surfaces use a proprietary anti-reflective coating to reduce signal loss to a minimum, achieving high optical transmission.

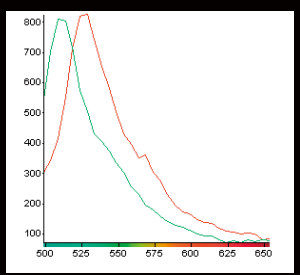
Accurate, reliable spectral data: three correction techniques

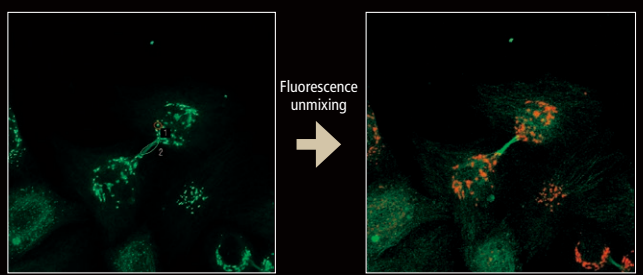
Three correction techniques allow for the acquisition of accurate spectra: interchannel sensitivity correction, which adjusts offset and sensitivity of each channel; spectral sensitivity correction, which adjusts diffraction grating spectral efficiency and detector spectral sensitivity; and correction of spectral transmission of optical devices in scan heads and microscopes.



Effortless fluorescence unmixing

Fluorescence labels with closely overlapping spectra can be unmixed cleanly with no crosstalk. Even without a given reference spectrum, simply specifying a Region of Interest (ROI) within the image and clicking the Simple Unmixing button allows separation of fluorescence spectra. The C2si+ contains a built-in database of given spectral data provided by manufacturers of fluorescence dyes that can be specified as reference spectra for fluorescence unmixing. Users may also add spectral information for new labels to the database.

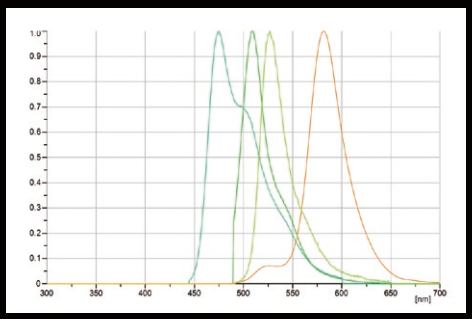


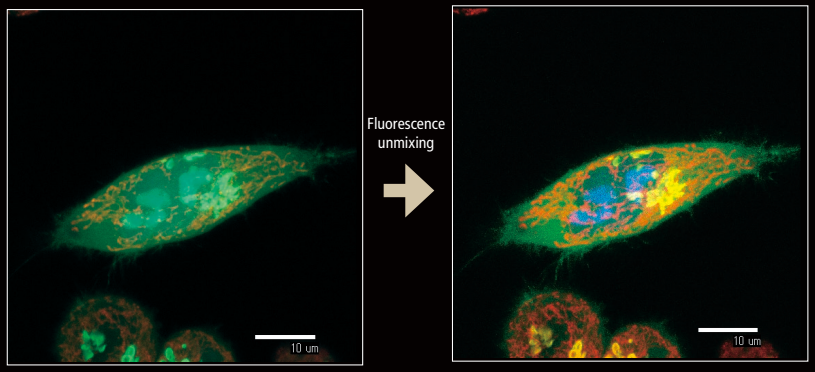


Specimen: HeLa cell in which GFP (Tubulin) and YFP (Golgi) are expressed. Spectral image captured with a 488 nm laser (left). After fluorescence unmixing, GFP is indicated in green and YFP is indicated in red (right). The graph (left) shows the spectral curve in the ROI.
Specimen courtesy of: Drs. Sheng-Chung Lee and Han-Yi E. Chou, National Taiwan University College of Medicine, Institute of Molecular Medicine

Unmixing of multiple fluorescence

Because wavelength resolution and range are freely selectable, scanning of a fluorescence protein with a wide wavelength range from blue to red such as CFP/GFP/YFP/DsRed is possible at one time. Reference data allows unmixing and display of each color.

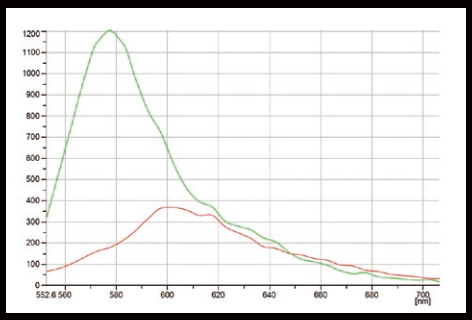




Specimen: HeLa cell in which nucleus is labeled with CFP, actin-related protein (Fascin) labeled with GFP, Golgi body labeled with YFP, and mitochondria labeled with DsRed. Spectral image captured with 408 nm and 488 nm laser exposure (left).
The fluorescence spectra of the captured image are unmixed using reference spectra (right).
Specimen courtesy of: Dr. Kaoru Katoh, The National Institute of Advanced Industrial Science and Technology (AIST)

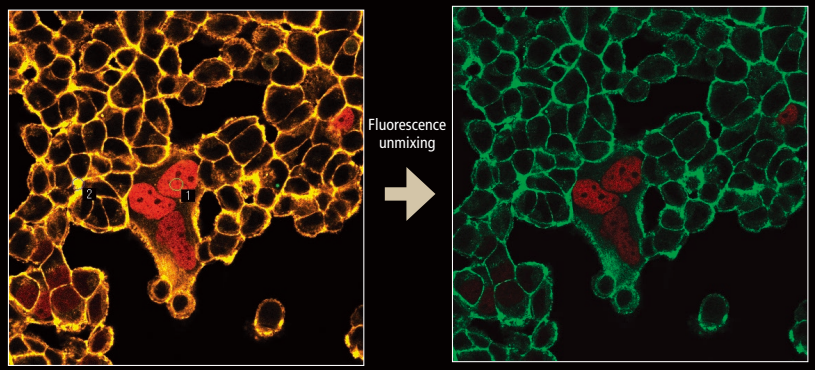
Unmixing red fluorochromes

Red fluorochromes, which had previously posed a challenge, are now simple to unmix.



Spectra for ROI 1 and 2 corresponding to the image on the right

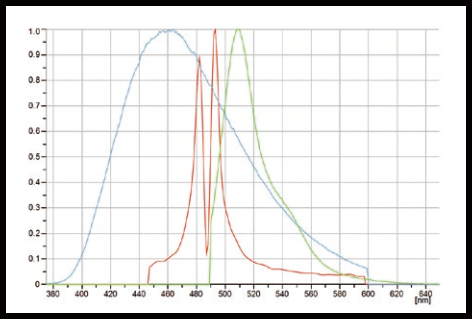
Rhodamine's fluorescence spectral peak is at approximately 579 nm, while that for RFP is approximately 600 nm. RFP's fluorescence is weaker than Rhodamine's, but their spectra are cleanly unmixed.

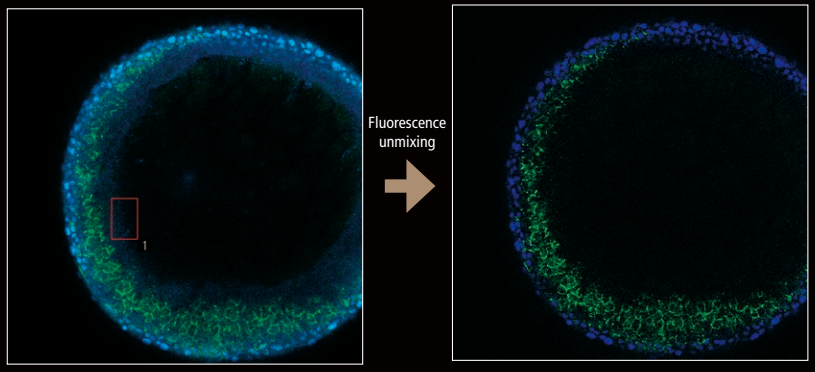


Specimen: actin of HeLa cell that has RFP expressed in the nucleus is stained with Rhodamine. Spectral image in the 550-630 nm wavelength range captured at 2.5 nm wavelength resolutions with 543 nm laser exposure (left). RFP indicated in red and Rhodamine indicated in green (right) in the image after fluorescence unmixing.

Unmixing auto-fluorescence of multi-stained samples

Fluorescence unmixing makes it possible not only to separate closely overlapping fluorescence spectra such as CFP and YFP but also to eliminate auto-fluorescence of cells, which until now was difficult.

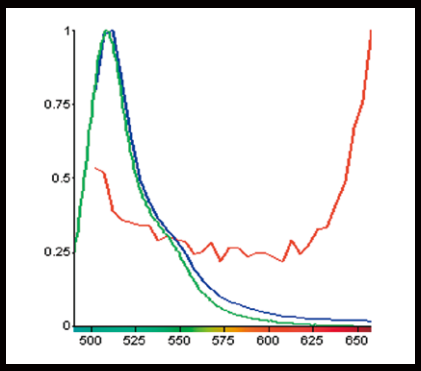




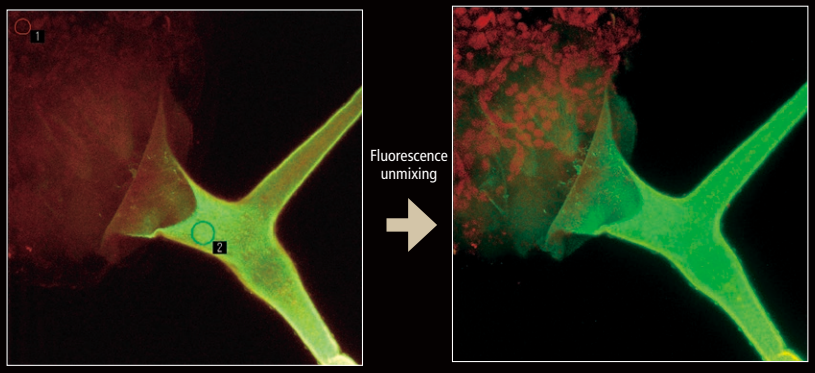
Specimen: Zebrafish egg stained with cadherin-GFP and DAPI. Spectral image captured with 408 nm and 488 nm laser exposures (left). After unmixing using reference spectra for auto-fluorescence (ROI1), GFP and DAPI, the auto-fluorescence in the image is eliminated (right).
Specimen courtesy of: Dr. Tohru Murakami, Neuromuscular and Developmental Anatomy, Gunma University Graduate School of Medicine

Confirmation of GFP expression

In conventional confocal observation, fluorescence is visualized as fluorescence intensity in a certain wavelength range. The spectral detector allows the confirmation of detailed wavelength characteristics of the fluorescence. The C2si$^{+/}$ spectral detector enables the slight color differences to be confirmed as wavelengths through sensitivity correction.



The correspondence of the spectral curve (blue) of ROI2 in the image and the reference curve (green) of eGFP proves that GFP is expressed in ROI2



Specimen: Arabidopsis proteoglycan and fused protein of GFP. Spectral image captured with 488 nm laser exposure (left). Once the image is unmixed using reference spectra for auto-fluorescence (ROI1) and GFP, GFP is indicated in green and auto-fluorescence is indicated in red (right).

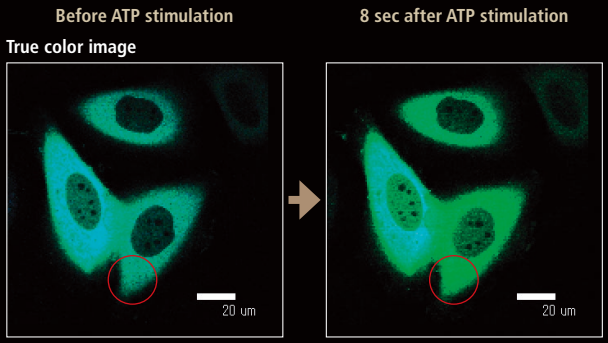
True spectral FRET analysis

FRET (Fluorescence Resonance Energy Transfer) analysis using true spectral imaging allows three-dimensional analysis with high signal-to-noise (S/N) ratio and high-spatial resolution as well as easy determination of FRET by real-time detection of spectral changes derived by FRET.

Also, even when spectra of donor and acceptor are overlapped like CFP and YFP, unmixing using reference data enables detection of detailed intensity changes and ratio analysis of fluorescence signals (YFP/CFP) without bleed through.

Acquisition of spectral image (XYT λ)

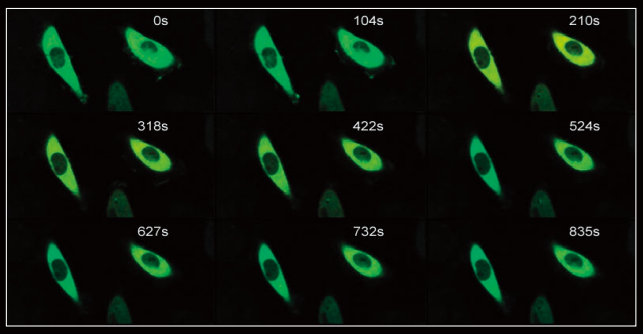
Spectral image in the 460-620 nm range captured at 5 nm wavelength resolution using a spectral detector enables observation of fluorescence wavelength changes.



True color image and spectral analysis of CFP and YFP. Spectral curve in ROI. Left peak indicates CFP and right peak indicates YFP respectively. After ATP stimulation, peak of CFP drops and peak of YFP rises due to FRET.

Fluorescence unmixing

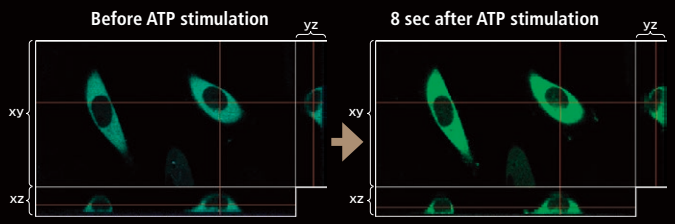
Spectral FRET analysis is possible by unmixing using reference data of CFP and YFP.
Two-dimensional change (FRET) of intracellular Ca^{2+} concentration is easily determined from spectral data without acceptor bleaching.



FRET image after spectral unmixing. CFP is indicated in blue and YPF indicated in green.

Five-dimensional analysis (XYZT λ)

Time-lapse changes (T) and spectra (λ) in three-dimensional space (XYZ) can be analyzed.



C2-DUVB GaAsP Detector Unit

The C2-DUVB is the first high-sensitivity GaAsP detector unit developed for the C2+ series. It is a compact fully tunable emission detector unit capable of spectral imaging with user-defined emission bandwidths in galvano imaging modalities.

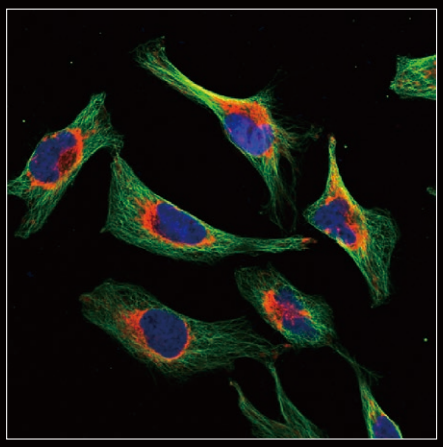
High-sensitivity spectral image acquisition

With a GaAsP PMT, the C2-DUVB tunable emission detector delivers flexible detection of fluorescent signals with higher sensitivity.

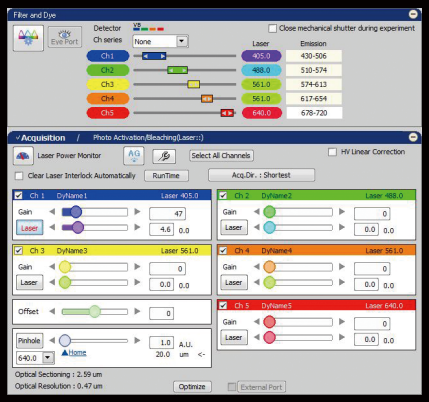
Variable acquisition wavelength range

User-defined emission bands can collect images within a selected wavelength range, replacing the need for fixed bandwidth emission filters.
Users can define the emission bandwidth range to as little as 10nm. Spectral images of multi-labeled specimens can be acquired by capturing a series of spectral images while changing detection wavelengths.

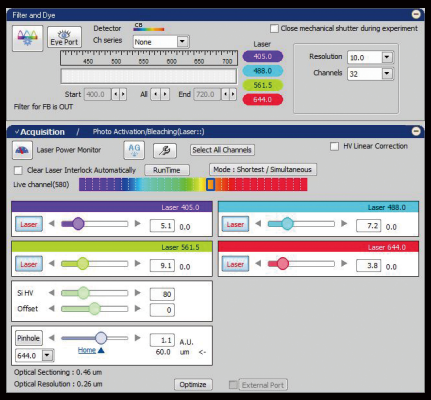
Based on the application, virtual bandpass mode and continuous bandpass modalities are selectable on the C2-DUVB.



VB (Variable Bandpass) mode
Captured in 3 ch



VB (Variable Bandpass) mode allows maximum 5ch color image



CB (Continuous Bandpass) allows maximum 32 ch spectrum imaging

Optional second channel detector

An optional second GaAsP PMT provides flexibility in detection. Users can divert selected wavelengths to the 2nd fixed bandwidth emission channel by inserting a dichroic mirror, while simultaneously utilizing the user-definable emission band on the first channel.
The second detector allows FRET, ratio imaging and other applications requiring simultaneous multi-channel imaging.

Accurate spectral unmixing

Multi-channel images acquired with the C2-DUVB can be spectrally unmixed by using the spectra of reference samples, or the spectra within the acquired images.

Laser Units

Two types of laser units are available for C2+/C2si+ confocal microscopes.

The compact model consisting of LU-N4, LU-N4S and LU-N3 comes pre-installed with either four (for LU-N4/4S) or three (LU-N3) lasers. The LU-N4/N4S/N3 model achieves maximal light throughput based on its alignment-free, high-efficiency laser combiner while maintaining a compact design. The flexible model, LU-NV, which also utilizes the high-efficiency laser combining system, supports up to eight different lasers and seven output fibers, an ideal laser system for multi-modal imaging.

LU-N4/N4S 4-laser unit, LU-N3 3-laser unit

The LU-N4/LU-N4S/LU-N3 model provides one output fiber. With its compact design, this laser unit provides a space-saving solution for confocal microscope systems.

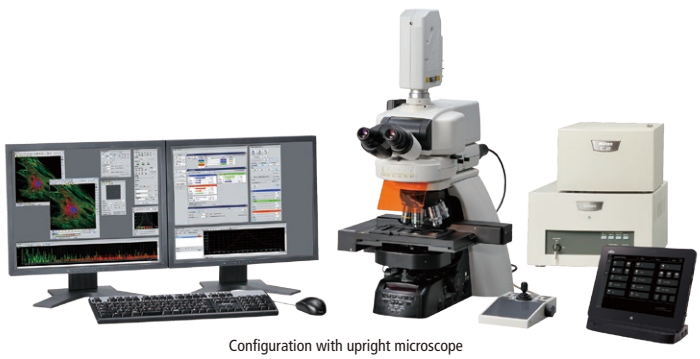
- The LU-N4/N3 is compatible with C2+, while LU-N4S is compatible with C2si+.
- The LU-N4/LU-N4S* is equipped with four lasers (405 nm, 488 nm, 561 nm, and 640 nm), while LU-N3 offers three lasers (405 nm, 488 nm, and 561 nm).
- The optical axis of each laser is aligned prior to shipment resulting in an effortless setup.
- The unique monolithic laser combining system prevents alignment shifts even after long-term use, resulting in consistent and maximal laser output.
- A built-in AOTF allows fast and efficient selection of wavelengths and power modulation.

LU-NV Series

The LU-NV laser combiner can be configured with up to eight different lasers and seven output fibers, providing maximal flexibility. With its multiple output fibers and fiber switching system, the LU-NV can support multiple laser applications, such as C2+/C2si+ confocal, TIRF and photoactivation on a single microscope platform.

- Available lasers for the LU-NV: 405 nm, 445 nm, 458 nm, 488 nm, 514 nm, 532 nm, 561 nm, 594 nm, 640 nm and 647 nm.
- High-power lasers for confocal microscopes are available.
- Lasers can be individually turned ON/OFF, boosting the efficiency of the lasers.
- The optical axis of each laser is aligned prior to shipment resulting in an effortless setup.
- The unique monolithic laser combining system prevents alignment shifts even after long-term use, resulting in consistent and maximal laser output.
- A built-in AOTF allows fast and efficient selection of wavelengths and power modulation.

LU-NV Laser Unit with LU Controller Box B (top)



Configuration with upright microscope

Configuration with inverted microscope

NIS-Elements High-performance Imaging Software

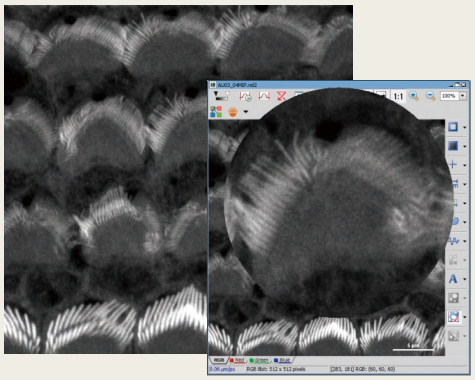
NIS-Elements C-ER

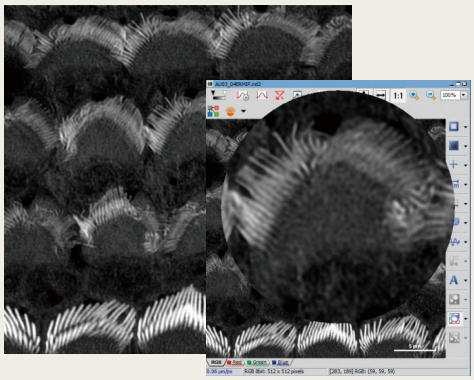
Automatic creation of high-resolution images

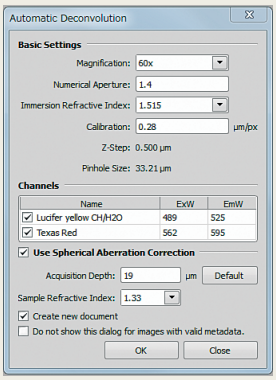
Higher resolution confocal images can be generated with a single click operation. The software assesses the captured image and automatically determines processing parameters to achieve enhanced resolution without sacrificing high imaging operability.
In addition, the resolution of previously captured confocal images can be improved using the pre-processing technology.

Improved resolution of confocal images

Image resolution is defined as the smallest distance between 2 points that can be resolved. The theoretical limit of resolution for a conventional optical microscope is approx. 200nm. Confocal microscopes potentially possess the capability of acquiring higher resolution images, but this has not been effectively achieved. With newly developed image processing technology, image resolution can be increased beyond that of a conventional confocal image (resolution can be improved 1.5 times (XY), 1.7 times (Z)).







Apical surfaces of auditory epithelia of mouse cochleae were stained by Atto-565-phalloidin at postnatal day 2.
Photographed with the cooperation of: Dr. Hideru Togashi, Division of Molecular and Cellular Biology,
Department of Biochemistry and Molecular Biology, Kobe University Graduate School of Medicine

* Use recommended objectives for confocal imaging when using NIS-Elements C-ER

NIS-Elements C-HC

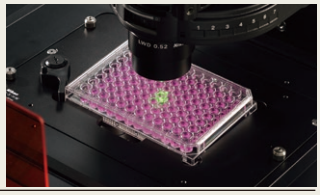
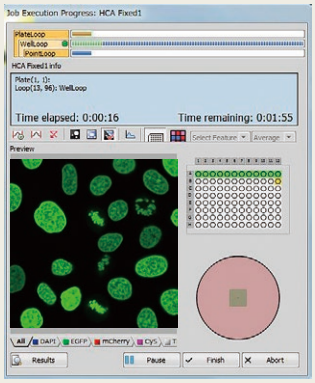


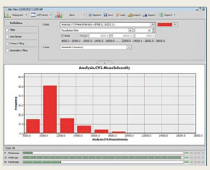
Image acquisition capability of large number of samples

Wizard format workflow simplifies settings for each experiment parameter. It enables flexible settings for the well plate, such as default registration of well plates from various manufacturers and definition of such XY capture point patterns as covering and random on the well.

Automated analysis during image acquisition

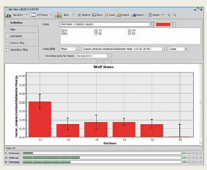
The current well-plate imaging point is displayed in real time along with a live image. Image analysis is conducted during image acquisition. Various formats are available for displaying results, including graphical display of analytical values such as histograms and scatter plots.



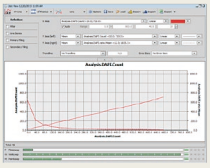


Histogram

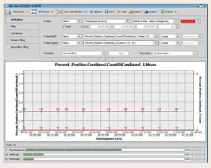
Scatter Plot



Bar Chart



XY Line



Time Chart

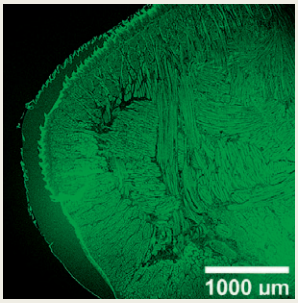
AZ-C2+ macro confocal microscope system

The AZ-C2+ macro confocal system enables confocal image acquisition of large specimens (> 1cm) with high resolution. Stunning, large field-of-view images with exceptionally high signal-to-noise ratios of whole embryos and large tissue slices can be captured in a single acquisition at pixel resolutions of up to 2048x2048.
Moreover, the AZ-C2+ macro confocal offers a combination of low and high magnification objective lenses, variable optical zoom and confocal scanning zoom function, enabling continuous imaging from macro to micro with a single microscope.
The AZ-C2+ macro confocal enables *in vivo* confocal imaging of macro specimens which is not possible with traditional stereoscopic microscopes.

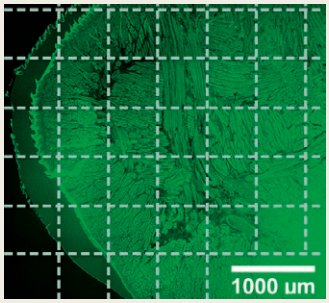
One-shot—whole specimen—macro confocal image acquisition

High NA objectives for macro observation enable fast, high-resolution, single-image capture of a wide specimen area. Because the objectives cover a field of view larger than 1 cm, imaging of embryos during late stages of development and the dynamics of cell populations in whole organs are possible. Minute specimen structures can be clearly seen, even in macro images.

Note: When Plan Apo 1x and AZ100 optical zoom 4x are used, the diagonal diameter of the real field of view is 5.3 mm.



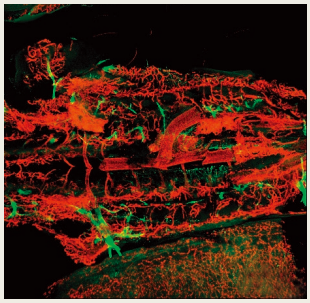
AZ-C2+ (Plan Fluor 2X used)



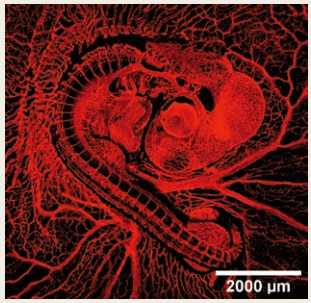
Conventional confocal microscope (stitched image)

With a conventional confocal microscope, image stitching is necessary because the field of view that can be captured in a single scan is small. The AZ-C2+ can capture a wide field of view—optical sections at high resolution in a single scan.

Specimen: Rat tongue slice

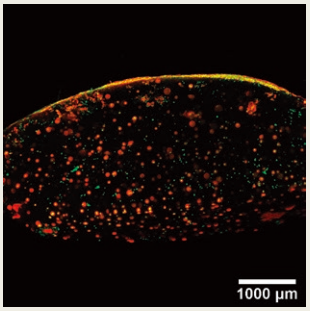


AZ-C2+ (Plan Apochromat 1X used)
Specimen: Neurons (green) and blood vessels (red) of 6.0-day-old chick embryo

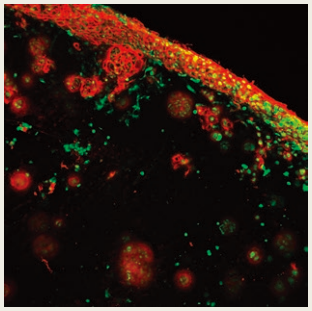


AZ-C2+ (Plan Apochromat 1X used)
Specimen: Blood vessels (red) of 2.5-dayold chick embryo

Photos courtesy of: Dr. Yoshiko Takahashi, Faculty of Science, Kyoto University

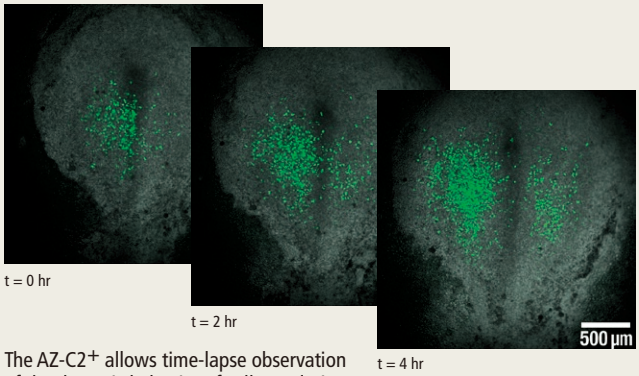


AZ-C2+ (macro image, Plan Apochromat 1X used)



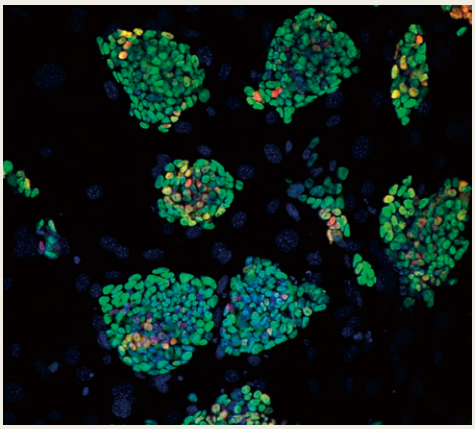
AZ-C2+ (magnified image)

Specimen: Rabbit hyaline cartilage cells embedded in atelocollagen gel and cultured for 21 days; live cells (green) and type II collagen (red)
Photos courtesy of: Dr. Masahiro Kino-oka, Laboratory of Bioprocess Systems Engineering, Department of Biotechnology, Division of Advanced Science and Biotechnology, Graduate School of Engineering, Osaka University



The AZ-C2+ allows time-lapse observation of the dynamic behavior of cell populations.

Specimen: Chick embryo in stage IV (Plan Fluor 2X used) expressing GAP43-eGFP to label plasma membrane.
Photos courtesy of: Dr. Yukiko Nakaya, Laboratory for Early Embryogenesis, Center for Developmental Biology, RIKEN



TT2 ES cells
Anti-Nanog antibody (Cy3), anti-Oct3/4 antibody (Alexa Fluor® 488) and DAPI localized in cell nuclei
Photo courtesy of: Dr. Hiroshi Kiyonari, Laboratory for Animal Resources and Genetic Engineering, RIKEN Center for Developmental Biology

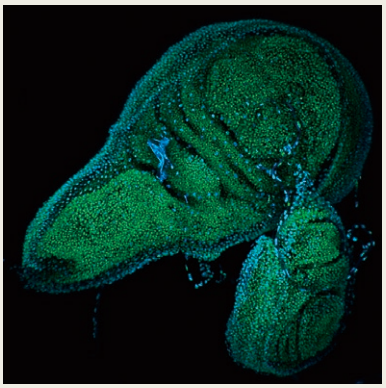
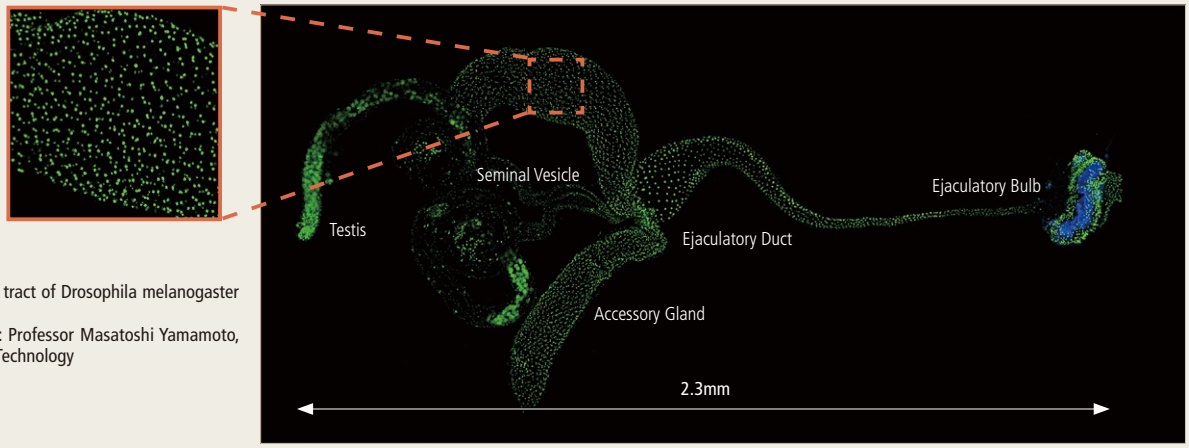


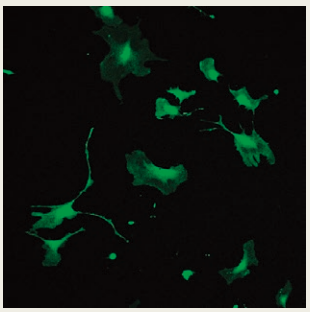
Photo courtesy of: Professor Masatoshi Yamamoto, Kyoto Institute of Technology



Specimen: Genital tract of Drosophila melanogaster (Stitched image)
Photo courtesy of: Professor Masatoshi Yamamoto, Kyoto Institute of Technology

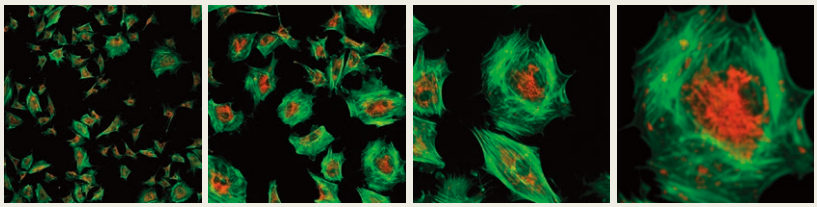
Continuous imaging from low magnification to high magnification

With five different objective lenses, optical zoom and confocal scan zoom, the AZ-C2$^+$ makes imaging possible from very low magnification to high magnification. Macro imaging, such as whole-section imaging, and micro imaging, including imaging of a single cell, can be done using a single microscope.



High magnification imaging offers clear and sharp images of single cells.

Specimen: Human breast cancer cell line MDA-MB-231 (Plan Fluor 5X used)
Photos courtesy of: Dr. Kazuyuki Itoh, Department of Biology, Research Institute, Osaka Medical Center for Cancer and Cardiovascular Diseases

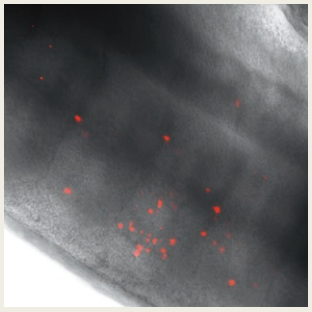


Zoom 1x | Zoom 2x | Zoom 4x | Zoom 8x

Specimen: BPAE cells (Plan Fluor 5X and C2$^+$ confocal scan zoom used)

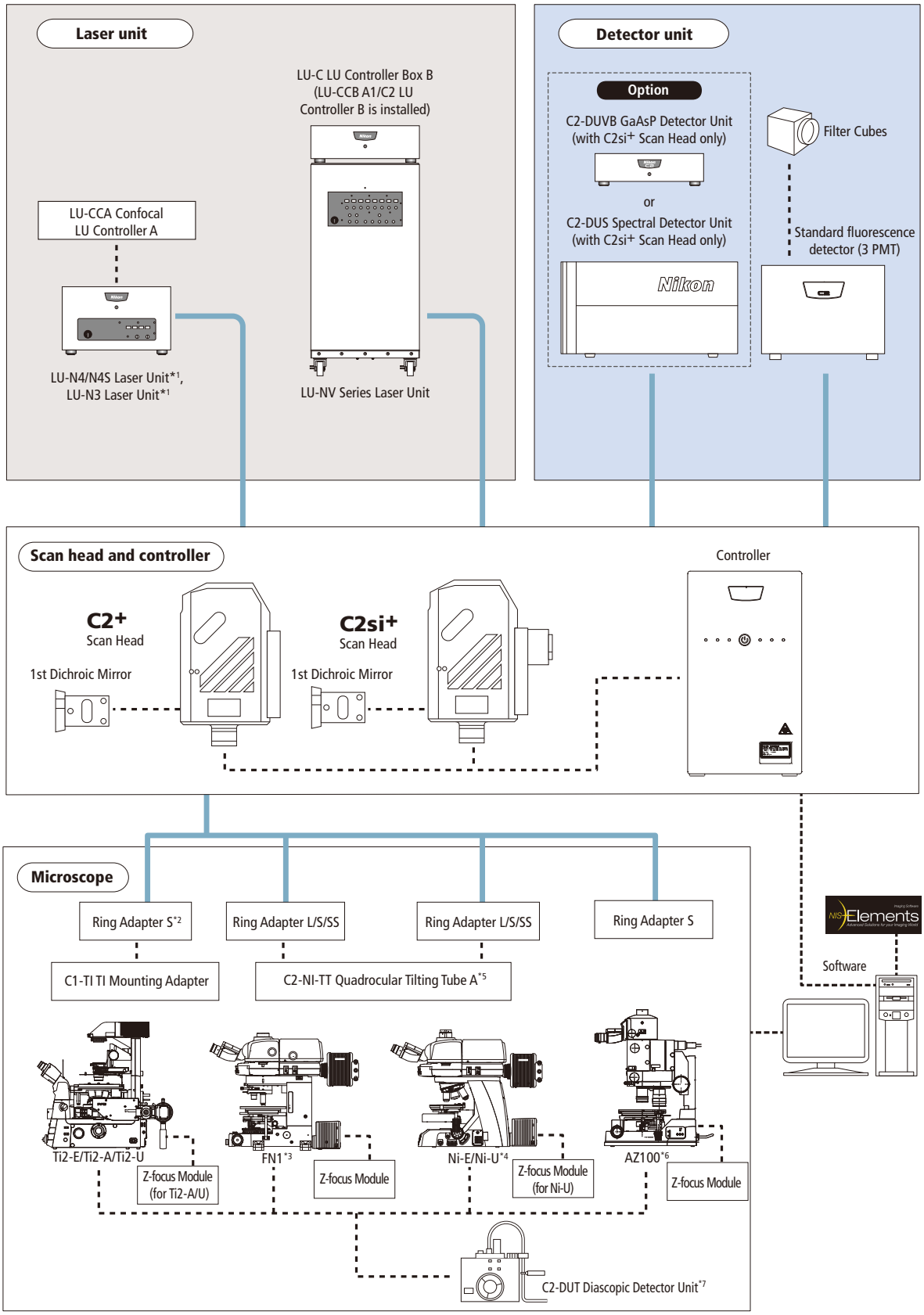
Deep imaging of whole specimens

The AZ-C2$^+$ allows imaging deep into the specimen—difficult to achieve with conventional confocal microscopes. The AZ-C2$^+$ efficiently captures fluorescence signals from deep within a specimen in both macro and micro imaging.



Observation with the AZ-C2$^+$
Cancer cells (red) 2 mm beneath the surface of the embryo can be imaged clearly.
Specimen: 2.5-day-old chick embryo
Photo courtesy of: Dr. Yoshiko Takahashi, Faculty of Science, Kyoto University

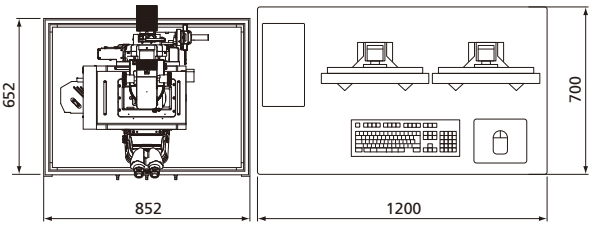
System diagram

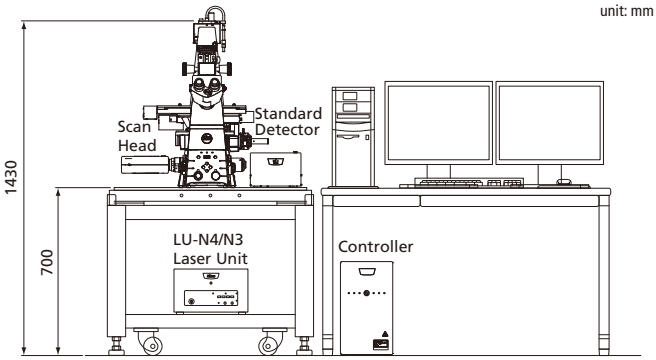


*1 LU-N4S is designed for use with the C2si+ spectral imaging confocal microscope. LU-N4/N3 is not compatible with C2si+.
*2 Not required when using a Stage-up Kit.
*3 C-TT-C Trinocular Tube or NI-TT Quadrocular Tilting tube can be used.
*4 With Ni-U, NI-TT Quadrocular Tilting tube or C-TT-C Trinocular Tube can be used.
With Ni-E, NI-TT-E Motorized Quadrocular Tilting tube, NI-TT Quadrocular Tilting tube or C-TT-C Trinocular Tube (for Ni-E focusing stage only) can be used.
*5 Required when using NI-TT Quadrocular Tilting tube or NI-TT-E Motorized Quadrocular Tilting tube.
*6 Use AZ-TE100LS Ergonomic Trinocular Tube 100 LS and AZ100 Stage Cover.
*7 Dedicated adapter is required depending on microscope model.

Recommended layout

unit: mm





Note) Computer table size is for reference only.

Specifications

	C2+
Laser unit	LU-N3: installed laser 405 nm, 488 nm, and 561 nm, Cannot be used with C2-DUS spectral detector LU-N4/N4S: installed laser 405 nm, 488 nm, 561 nm, and 640 nm, Use LU-N4S when using C2-DUS spectral detector LU-NV series: mountable laser 405 nm, 445 nm, 458 nm, 488 nm, 514 nm, 532 nm, 561 nm, 594 nm, 640 nm and 647 nm
Standard detector	Wavelength: 400-750 nm, Detector: 3PMTs, Filter cube: 2 filter cubes
Diascopic detector (option)	Wavelength: 400-700 nm, Detector: 1 PMT
Scan head	C2+/C2si+ scan head: when configuring with C2-DU3 standard fluorescence detector Scanner: galvano scanner, Pixel size: 2048 x 2048 Scanning speed: Standard mode; 2 fps (512 x 512 pixels, bi-direction), 18 fps (512 x 32 pixels, bi-direction), Zoom: 1-1000x Fast mode; 8 fps (512 x 512 pixels, bi-direction), 100 fps (512 x 32 pixels, bi-direction)*1, Zoom: 8-1000x
	C2si+ scan head: when configuring with C2-DUS spectral detector Scanner: galvano scanner, Pixel size: max. 1024 x 1024 pixels Scanning speed: 0.5 fps (512 x 512 pixels, single direction), 7 fps (512 x 32 pixels, single direction)
Scan mode	X-T, Y-T, X-Y, X-Y-T, X-Y-Z, X-Y-Z-T, Mirror Image (X, Y), Reciprocation image, Rotation image, ROI image, Line sequential acquisition
Pinhole	Circular shape, 6 size
Spectral detector (option)*2	C2-DUS spectral detector unit Number of channels: 32, Wavelength detection range: 400 - 750 nm, Spectral image acquisition speed: 2 fps (256 x 256 pixels) Maximum pixel size: 1024 x 1024 (Spectral mode/Virtual filter mode), Wavelength resolution: 2.5/5.0/10.0 nm, Wavelength range variable in 0.25 nm steps
	C2-DUVB GaAsP detector unit Number of channels: 1 GaAsP PMT with variable emission plus 1 optional GaAsP PMT (C2-DUVB-OP) with a user-defined dichroic mirror and barrier filter, Wavelength detection range: 400 - 720 nm, narrowest: 10 nm, broadest: 320 nm, Maximum pixel size: 2048 x 2048 (CB mode/VB mode), Wavelength resolution: 10 nm, Wavelength range variable in 1 nm steps
FOV	Square inscribed in a ø18 mm circle
Image bit depth	12 bits
Compatible microscopes	Inverted microscope ECLIPSE Ti2-E/Ti2-A/Ti2-U, Upright microscope ECLIPSE Ni-E (focusing nosepiece type/focusing stage type)/Ni-U, Fixed stage microscope ECLIPSE FN1, Multi-purpose zoom microscope AZ100
Z step	Ti2-E: 0.02 µm, FN1 stepping motor: 0.05 µm, Ni-E: 0.025 µm
Software	Software: NIS-Elements C Display/image processing/analysis 2D/3D/4D analysis, time-lapse analysis, 3D volume rendering/orthogonal, spatial filters, image stitching, multipoint time-lapse, spectral unmixing, real-time unmixing, virtual filters, deconvolution, AVI image file output
	Application: FRAP, FLIP, FRET, photoactivation, colocalization, three-dimensional time-lapse imaging, multipoint time-lapse imaging
Control Computer	OS: Microsoft Windows 7 Professional 64 bit SP1, Windows 10 Pro 64 bit, CPU: Intel Xeon Duo Processor E5-1630v3 (3.70 GHz/10 MB/2133 MHz/Quad Core) or higher, RAM: DDR4 2400MHz 8GB, Chipset: Intel C612 chipset, HDD: Serial ATA 6 Gb/s (7,200 rpm) 146 GB or greater, Graphics: NVIDIA Quadro M2000 4GB, Drive: Super Multi drive (Slimline), Expansion slots: PCI Express (x16) (for graphics) slot x2 etc., Interface: USB2.0 RS-232C, LAN port: 10/100/1000 network interface x1, Monitor: LCD2 monitor with 1600 x 1200 or higher resolution, dual monitor, Configuration is recommended
Dimensions and weight	C2+/C2si+ scan head 145 (W) x 236.5 (H) x 70 (D) mm, Approx. 2 kg Controller 225 (W) x 404 (H) x 382 (D) mm, Approx. 12 kg Standard detector 225 (W) x 154 (H) x 224 (D) mm, Approx. 3.5 kg Spectral detector C2-DUS: 289 (W) x 323 (H) x 593.5 (D) mm, Approx. 23 kg, C2-DUVB: 360 (W) x 91 (H) x 595.5 (D) mm, Approx. 9.5 kg

*1 The described frame rate is NOT available with Rotation, CROP, ROI, Spectral imaging and Stimulation.
*2 Only available with C2si+ scan head.

Specifications and equipment are subject to change without any notice or obligation on the part of the manufacturer. June 2017 ©2010-17 NIKON CORPORATION

⚠ WARNING	TO ENSURE CORRECT USAGE, READ THE CORRESPONDING MANUALS CAREFULLY BEFORE USING YOUR EQUIPMENT.

Monitor images are simulated.
Some sample images in this brochure were captured using the C1 confocal microscope system.
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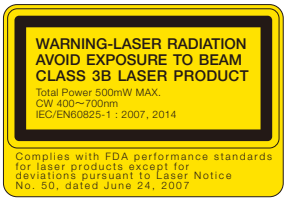
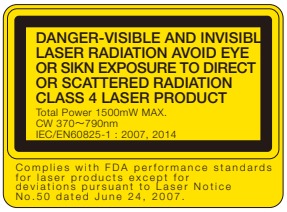
*Products: Hardware and its technical information (including software)





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